



Technical Manual

PHE A (Phenylethanolamine A) ELISA Kit

- **Catalogue Code: FSES0012**
- **Competitive ELISA Kit**
- **Research Use Only**

Contents

1. Key features and Sample Types	3
2. Storage	3
3. Test Principle.....	3
4. Kit Contents.....	4
5. Experimental Preparation	5
6. Assay Procedure.....	6
7. Data Analysis.....	7
8. Notes.....	7

1. Key features and Sample Types

Sensitivity:

0.1 ppb (ng/mL)

Assay Procedure:

25°C, 30 min~15 min

Detection Limit:

Urine - 0.1 ppb; Muscle - 0.1 ppb; Feed - 1 ppb

Cross Reactivity:

PHE A - 100%, Clenbuterol - < 1%, Salbutamol - < 1%, Ractopamine - < 1%

Sample Recovery rate:

Urine - 95%±15%, Muscle, Feed - 85%±15%,

Storage:

2-8°C for 6 months.

Expiry:

See Kit Label

2. Storage

Store the kit at 2~8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2 - 8°C.

3. Test Principle

This kit uses a Competitive-ELISA method. It can detect Phenylethanolamine A (PHE A) in samples, such as muscle, feed. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, PHE A in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-PHE A antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of PHE A. The concentration of PHE A in the samples can be calculated by comparing the OD of the samples to the standard curve.

4. Kit Contents

Each kit contains reagents for 96 assays including:

No.	Component	96-WellKit
1	ELISA Microtiter plate	96 wells
2	Standards	1 mL each (0ppb, 0.1ppb, 0.3ppb, 0.9ppb, 2.7ppb, 8.1ppb)
3	HRP Conjugate	5.5 mL
4	Antibody Working Solution	5.5 mL
5	Substrate Reagent A	6 mL
6	Substrate Reagent B	6 mL
7	Stop Solution	6 mL
8	20xConcentrated Wash Buffer	40 mL
9	10xReconstitution Buffer	50 mL
10	Plate Sealer	3 pieces
11	Sealed Bag	1 piece
12	Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Additional materials required:

Other materials required but not supplied

- **Instruments:** Microplate reader, Printer, Homogenizer, Nitrogen evaporators, Water bath, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).
- **Micropipette:** Single channel (20-200 μ L, 100-1000 μ L), Multichannel (30-300 μ L).
- **Reagents:** NaOH, Ethyl acetate, Concentrated HCl, Acetonitrile, Methanol, N-hexane, Na₂SO₄.

5. Experimental Preparation

Bring all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

1. Sample pre-treatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross- contamination during the experiment.

2. Solution preparation

Solution 1: 0.1 M HCl Solution

Dilute 0.86 mL of **Concentrated HCl** to 100mL with deionized water.

Solution 2: 0.1 M NaOH Solution (*for muscle sample*)

Dissolve 0.4 g of **NaOH** to 100mL with deionized water.

Solution 3: Acetonitrile- **0.1 M HCl Solution** (Solution 1)

Acetonitrile (V): 0.1 M HCl (V) =84:16 (*for tissue sample*)

Solution 4: Reconstitution Buffer (*for muscle, feed sample*)

Dilute the **10×Reconstitution Buffer** with deionized water. (10×Reconstitution Buffer (V): Deionized water (V)=1:9). The Reconstitution buffer can be store at 4°C for a month.

Solution 5: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water. (20×Concentrated Wash Buffer (V): Deionized water (V) = 1:19).

3. Sample pre-treatment procedure

Substance in sample is distributed unevenly. It is recommended that more samples should be taken when sampling.

Targets may be distributed unevenly, resulting in no detection. To avoid this, ensure to take sufficient samples when sampling.

3.1 Pre-treatment of urine sample:

1. Take 50 μ L of clear urine sample to detect (if the urine sample is turbid, it should be filtered or centrifuged at 4000 r/min for 5 min until the urine sample become clear). Temporarily used samples should be kept frozen to save.

Note: Sample dilution factor: 1, detection limit: 0.1ppb

3.2 Pre-treatment of muscle sample:

1. Weigh 2 ± 0.05 g of homogenate muscle sample, add 6mL of **Acetonitrile - 0.1 M HCl Solution** (Solution 3), vortex for 2 min, and centrifuge at 4000 r/min at room temperature for 10 min.
2. Take 3 mL of the supernatant, add 2 mL of **0.1 M NaOH Solution** (Solution 2), add 6 mL of **Ethyl acetate**, vortex for 2 min, and centrifuge at 4000 r/min at room temperature for 10 min. Take all the supernatant to dry at 50-60°C with nitrogen evaporators or water bath.
3. Add 1 mL of **Reconstitution Buffer** (Solution 4), mix and vortex for 30s, take 50

uL liquid for analysis

Note: Sample dilution factor: 1, detection limit: 0.1ppb

3.3 Pre-treatment of feed sample:

1. Weigh 1.0 ± 0.05 g of homogenate feed sample, add 10 mL of **Methanol**, then add 5 g of **Na₂SO₄**, vortex for 2 min, and centrifuge at 4000 r/min at room temperature for 10min.
2. Take 1 mL of supernatant, dry at 50-60°C with nitrogen evaporators or water bath. Dissolve the dried residual with 1 mL of **Reconstitution Buffer (Solution 4)**, add 1 mL of **N-hexane** and mix for 30s. Centrifuge at 4000 r/min at room temperature for 5 min.
3. Take 50 µL of lower liquid for analysis.

Note: Sample dilution factor: 10, detection limit: 1ppb

6. Assay Procedure

Bring all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2~8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples must be tested in duplicate.**
2. **Add Sample:** add 50 µL of **Standard or Sample** per well, then add 50 µL of **HRP Conjugate** to each well, then add 50 µL of **Antibody Working Solution**, cover the plate with plate sealer. Vortex for 5s gently to mix thoroughly, incubate at 25°C for 30 min away from direct sunlight.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 300 µL of **Wash Buffer (Solution 5)** to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Colour Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently vortex for 5s to mix thoroughly. Incubate away from direct sunlight for 15 min at 25°C.
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, vortex gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

7. Data Analysis

1. Absorbance (%) = $A/A_0 \times 100\%$

A: Average absorbance of standard or sample

A_0 : Average absorbance of 0 ppb Standard

2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration.

If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

8. Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microplate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the FSES0012. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other FSES0012 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue colour. When OD value of standard (concentration: 0) < 0.5 unit ($A_{450nm} < 0.5$), it indicates the reagents are deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Assay Genie 100% money-back guarantee!

If you are not satisfied with the quality of our products and our technical team cannot resolve your problem, we will give you 100% of your money back.

Contact Details



Email: info@AsssayGenie.com

Web: www.AsssayGenie.com