



Technical Manual

SQX (Sulfaquinoxaline) ELISA Kit

- Catalogue Code: FSES0034
- Competitive ELISA Kit
- Research Use Only

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1. Key features and Sample Types

Sensitivity:

0.5 ppb (ng/mL)

Assay Procedure:

25°C, 45 min~ 15 min

Detection Limit:

Muscle (method 1) - 0.5 ppb; Muscle (method 2) - 2.5 ppb;

Serum, Urine - 2 ppb; Honey - 0.5 ppb; Milk - 10 ppb; Feed - 20 ppb.

Cross Reactivity:

Names	Cross-reactivity
Sulfaquinoxaline(SQX)	100%
Sulfadiazine (SD or SDZ)	<1%
Sulfamerazine(SM1)	<1%
Sulfamethazine (SM2)	<1%
Sulfamonomethoxine(SMM)	<1%

Sample Recovery rate:

Muscle, Honey - 95%±25%; Urine, Milk, Serum, Feed - 85%±25%

Storage:

2-8°C for 6 months.

Expiry:

See Kit Label

2. Storage

Store the kit at 2~8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2 - 8°C.

3. Test Principle

This kit uses a Competitive-ELISA method. It can detect Sulfaquinoxaline (SQX) in samples, such as muscle, milk, honey, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, SQX in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-SQX antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and TMB substrate is added for color

development. There is a negative correlation between the OD value of samples and the concentration of SQX. The concentration of SQX in the samples can be calculated by comparing the OD of the samples to the standard curve.

4. Kit Contents

Each kit contains reagents for 96 assays including:

No.	Component	96-WellKit
1	ELISA Microtiter plate	96 wells
2	Standards	1 mL each (0 ppb, 0.5 ppb, 1.5 ppb, 4.5 ppb, 13.5 ppb, 40.5 ppb)
3	HRP Conjugate	5.5 mL
4	Antibody Working Solution	5.5 mL
5	Substrate Reagent A	6 mL
6	Substrate Reagent B	6 mL
7	Stop Solution	6 mL
8	20×Concentrated Wash Buffer	40 mL
9	2×Reconstitution Buffer	50 mL
10	Plate Sealer	3 pieces
11	Sealed Bag	1 piece
12	Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Additional materials required:

Other materials required but not supplied

- **Instruments:** Microplate reader, Printer, Homogenizer, Nitrogen evaporators, Water bath, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01g).
- **Micropipette:** Single channel (20-200 µL, 100-1000 µL), Multichannel (30-300 µL).
- **Reagents:** Ethyl acetate, N-hexane, Acetonitrile, $\text{Na}_2\text{HPO}_4 \cdot 12\text{H}_2\text{O}$, NaOH, Concentrated HCl, $\text{NaH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$.

5. Experimental Preparation

Bring all reagents and samples to room temperature before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters.

1. Sample pre-treatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross- contamination during the experiment.

2. Solution preparation

Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

Solution 1: 0.2 M NaOH Solution (for honey sample)

Dissolve 0.8 g of **NaOH** to 100 mL with deionized water.

Solution 2: 0.02 M PB Buffer (for muscle, serum, urine, milk, feed sample)

Dissolve 2.58 g of **Na₂HPO₄·12H₂O** and 0.44 g of **NaH₂PO₄·2H₂O** to 500 mL with deionized water.

Solution 3: 0.5 M HCl Solution (for honey sample)

Add 4.3 mL of **Concentrated HCl** to 100mL with deionized water and mix thoroughly.

Solution 4: Reconstitution Buffer (for muscle, honey sample)

Dilute the **2×Reconstitution Buffer** with deionized water. (2×Reconstitution Buffer (V): Deionized water (V)=1:1). The reconstitution buffer can be store at 4°C for a month.

Solution 5: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water.

(20×Concentrated Wash Buffer (V): Deionized water (V) = 1:19).

3. Sample pre-treatment procedure

Targets may be distributed unevenly, resulting in no detection. To avoid this, ensure to take sufficient samples when sampling.

3.1 Pre-treatment of muscle (method 1) sample:

1. Add 3±0.05 g of homogeneous muscle sample to a centrifuge tube, then add 3 mL of **0.02 M PB Buffer** (Solution 2), vortex and mix thoroughly. Add 4 ml of **Ethyl acetate** and 2 mL of **Acetonitrile**, vortex for 10 min, centrifuge at 4000 r/min for 10 min.
2. Take 2 mL of upper liquid, dry at 50-60°C with nitrogen evaporators or water bath.
3. Dissolve the residual with 1 mL of **N-hexane**, add 1 mL of **Reconstitution Buffer** (Solution 4) and vortex for 1 min. Centrifuge at 4000 r/min for 5 min.
4. Discard the upper n-hexane, take 50 µL of lower liquid for analysis.

Note: Sample dilution factor: 1, minimum detection limit: 0.5 ppb

3.2 Pre-treatment of muscle (method 2) sample:

1. Add 2.0 ± 0.05 g of homogeneous muscle sample to a centrifuge tube, then add 8 mL of **0.02 M PB Buffer** (Solution 2), vortex for 2 min, centrifuge at 4000 r/min for 10 min.
2. Take 50 μ L of liquid for analysis.

Note: Sample dilution factor: 5, minimum detection limit: 2.5 ppb

3.3 Pre-treatment of serum (swine)* sample:

(*Data validated in swine serum but pre-treatment can be applied for serum samples of multiple species.)

1. Stand blood sample at room temperature for 30 min, centrifuge at 4000 r/min for 10 min, separate the serum.
2. Take 1 mL of serum, add 3 mL of **0.02 M PB Buffer** (Solution 2), and mix for 30s.
3. Take 50 μ L of liquid for analysis.

Note: Sample dilution factor: 4, minimum detection limit: 2 ppb

3.4 Pre-treatment of honey sample:

1. Weigh 1 ± 0.05 g of honey sample into 50 mL a centrifuge tube, add 1 mL of **0.5 M HCl Solution** (Solution 3), incubate at 37°C for 30 min.
2. Add 2.5 mL of **0.2 M NaOH solution** (Solution 1) (adjust the PH value to about 5), then add 4 mL of **Ethyl acetate**, vortex for 5 min, centrifuge at 4000 r/min at room temperature for 10 min.
3. Take 2 mL of upper liquid, dry at 50-60°C with nitrogen evaporators or water bath. Add 0.5 mL of **Reconstitution Buffer** (Solution 4) and mix for 30s.
4. Take 50 μ L of liquid for analysis.

Note: Sample dilution factor: 1, minimum detection limit: 0.5 ppb

3.5 Pre-treatment of urine (swine)* sample:

(*Data validated in swine urine but pre-treatment can be applied for urine samples of multiple species.)

1. Mix 3 mL of **0.02 M PB Buffer** (Solution 2) and 1 mL of centrifuged clear urine sample for 30s.
2. Take 50 μ L of liquid to analysis.

Note: Sample dilution factor: 4, minimum detection limit: 2 ppb

3.6 Pre-treatment of milk sample:

1. Dilute milk sample with **0.02 M PB Buffer** (Solution 2) (for example, 20 μ L milk + 380 μ L of 0.02 M PB Buffer (Solution 2)), mix for 30s.
2. Take 50 μ L of liquid to analysis.

Note: Sample dilution factor: 20, minimum detection limit: 10 ppb

3.7 Pre-treatment of feed sample:

1. Weigh 2 g of crushed homogenate into the 50 mL centrifuge tube, add 8 mL of **Acetonitrile**, vortex for 5 min, centrifuge at 4000 r/min for 5 min at room temperature.
2. Take 1 mL of upper organic phase to glass tube and dry with nitrogen evaporators/water bath at 50-60°C.
3. Add 1 mL of **N-hexane**, vortex for 30s. Then add 1 mL of **0.02M PB Buffer** (Solution 2), vortex for 30s, centrifuge at 4000 r/min for 5 min at room temperature.

4. Remove the upper organic phase, take 100 µL of upper water phase to another tube. Add 900 µL of **0.02M PB Buffer** (Solution 2), vortex sufficiently for 5 min, mix fully;
5. Take 50 µL for detection and analysis.

Note: Sample dilution factor: 40, minimum detection limit: 20 ppb

6. Assay Procedure

Bring all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2~8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples must be tested in duplicate.**
2. **Add Sample:** add 50 µL of **Standard or Sample** per well, then add 50µL of **HRP Conjugate** to each well, then add 50 µL of **Antibody Working Solution**, cover the plate with plate sealer, vortex for 5s gently to mix thoroughly, incubate at 25°C for 45 min away from direct sunlight.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 300 µL of **Wash Buffer** (Solution 5) to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Colour Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently vortex for 5s to mix thoroughly. Incubate at 25°C for 15 min away from direct sunlight (The reaction time can be extended according to the actual colour change).
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, vortex gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

7. Data Analysis

1. **Absorbance (%) = $A/A_0 \times 100\%$**

A: Average absorbance of standard or sample

A₀: Average absorbance of 0 ppb Standard

2. **Drawing and calculation of standard curve**

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

8. Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microplate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the FSES0034. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other FSES0034 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue colour. When OD value of standard (concentration: 0) < 0.5 unit (A450nm < 0.5), it indicates the reagents are deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

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