



Technical Manual

TMP (Trimethoprim) ELISA Kit

- Catalogue Code: FSES0068
- Competitive ELISA Kit
- Research Use Only

Contents

1. Key features and Sample Types	3
2. Storage	3
3. Test Principle.....	3
4. Kit Contents.....	4
5. Experimental Preparation	5
6. Assay Procedure.....	6
7. Data Analysis	6
8. Notes	7

1. Key features and Sample Types

Sensitivity:

0.1 ppb (ng/mL)

Assay Procedure:

25°C, 30min~15min

Detection Limit:

Meat - 10ppb

Cross Reactivity:

Trimethoprim - 100%

Sample Recovery rate:

90%±30%.

Storage:

2-8°C for 6 months.

Expiry:

See Kit Label

2. Storage

Store the kit at 2~8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2~8°C.

3. Test Principle

This kit uses a Competitive-ELISA method. It can detect Trimethoprim (TMP) in samples, such as raw milk, muscle, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, TMP in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-TMP antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of TMP. The concentration of TMP in the samples can be calculated by comparing the OD of the samples to the standard curve.

4. Kit Contents

Each kit contains reagents for 96 assays including:

No.	Component	96-WellKit
1	ELISA Microtiter plate	96 wells
2	Standards	1 mL each (0ppb, 0.1ppb, 0.3ppb, 0.9ppb, 2.7ppb, 8.1ppb)
3	HRP Conjugate	7mL
4	Antibody Working Solution	7mL
5	8×Concentrated Sample Solution	25mL
6	Sample Diluent	7mL
7	Substrate Reagent A	6mL
8	Substrate Reagent B	6mL
9	Stop Solution	6mL
10	20×Concentrated Wash Buffer	25mL
11	Plate Sealer	3 pieces
12	Sealed Bag	1 piece
13	Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Additional materials required:

Other materials required but not supplied

- **Instruments:** Microplate reader, Homogenizer, Vortex mixer, Nitrogen evaporators, Water bath, Centrifuge, Balance (sensitivity 0.01 g).
- **Micropipette:** Single channel (20-200 μ L, 100-1000 μ L), Multichannel (30-300 μ L).
- **Reagents:** N-hexane, Acetonitrile

5. Experimental Preparation

Bring all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

1. Sample pre-treatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross- contamination during the experiment.

2. Solution preparation

Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

Solution 1: Sample Solution

Dilute **8xConcentrated Sample Solution** with deionized water.

(8xConcentrated Sample Solution (V): Deionized water (V) = 1:7). Mix fully.

Solution 2: Wash Buffer

Dilute **20xConcentrated Wash Buffer** with deionized water

(20xConcentrated Wash Buffer (V): Deionized water (V) = 1:19). Mix fully.

3. Sample pre-treatment procedure

Targets may be distributed unevenly, resulting in no detection. To avoid this, ensure to take sufficient samples when sampling.

3.1 Pre-treatment of muscle (pork, chicken, duck) sample:

1. Weigh 3 ± 0.05 g of homogenate sample into 50 mL centrifuge tub. Add 50 μ L of **Sample Diluent** and 6 mL of **Acetonitrile**, vortex for 2 min, mix fully. Centrifuge at 4000 r/min for 5 min.
2. Take 3 mL of supernatant to another centrifuge tube and add dry at 50-60°C with nitrogen evaporators or water bath.
3. Add 2 mL of **N-hexane** to dissolve the remaining dry material, then add 1 mL of **Sample Solution** (Solution 1). Vortex for 1 min and Centrifuge at 4000 r/min for 5 min.
4. Remove the upper layer n-hexane phase and intermediate layer.
5. Take 50 μ L of the lower layer liquid to another centrifuge tube, add 450 μ L of **Sample Solution** (Solution 1).
6. Take 20 μ L of liquid for analysis.

Note: Sample dilution factor: 10, detection limit: 10ppb.

6. Assay Procedure

Bring all reagents and samples to room temperature before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2~8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples must be tested in duplicate.**
2. **Add Sample:** add 20µL of **Standard or Sample** per well, then add 50 µL of **HRP Conjugate** and add 50µL of **Antibody Working Solution** to each well, cover the plate with plate sealer. Vortex for 10s gently to mix thoroughly. Incubate at 25°C for 30 min away from direct sunlight.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 260 µL of **Wash Buffer** (Solution 2) to each well and wash. Repeat wash procedure for 4 times, 30s intervals/time. Invert the plate and pat it against with absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Colour Development:** add 50µL of **Substrate Reagent A** to each well, and then add 50µL of **Substrate Reagent B**. Gently vortex for 10s to mix thoroughly. Incubate at 25°C for 15 min away from direct sunlight (The reaction time can be extended according to the actual colour change).
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well. Gently vortex for 10s to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 5 min after stop reaction.

7. Data Analysis

1. **Absorbance (%) = $A/A_0 \times 100\%$**

A: Average absorbance of standard solution or sample

A₀: Average absorbance of 0 ppb Standard solution

2. **Drawing and calculation of standard curve**

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add the average absorbance value to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

8. Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the FSES0068. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other FSES0068 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue colour. When OD value of standard (concentration: 0) < 0.8 unit (A450nm < 0.8), it indicates the reagents are deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Assay Genie 100% money-back guarantee!

If you are not satisfied with the quality of our products and our technical team cannot resolve your problem, we will give you 100% of your money back.

Contact Details



Email: info@AsssayGenie.com

Web: www.AssayGenie.com