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## Technical Manual

### FB1 (Fumonisin B1) ELISA Kit

- Catalogue Code: FSES0079
- Competitive ELISA Kit
- Research Use Only

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## 1. Key features and Sample Types

### Sensitivity:

1 ppb (ng/mL)

### Assay Procedure:

25°C, 30 min~15 min

### Detection Limit:

Corn, Feed - 150 ppb; Edible oil - 20 ppb

### Cross Reactivity:

Fumonisin B1 (FB1) - 100%

### Sample Recovery rate:

Feed - 95%±15%; Corn - 100%±15%; Edible oil - 85%±15%

### Storage:

2-8°C for 6 months.

### Expiry:

See Kit Label

## 2. Storage

Store the kit at 2~8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2 - 8°C.

## 3. Test Principle

This kit uses a Competitive-ELISA method. It can detect Fumonisin B1 (FB1) in samples, such as corn, feed, edible oil, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, FB1 in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-FB1 antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of FB1. The concentration of FB1 in the samples can be calculated by comparing the OD of the samples to the standard curve.

## 4. Kit Contents

Each kit contains reagents for 96 assays including:

No.	Component	96-WellKit
1	ELISA Microtiter plate	96 wells
2	Standards	1mL each (0 ppb, 1 ppb, 3 ppb, 9 ppb, 27 ppb, 81 ppb)
3	HRP Conjugate	5.5 mL
4	Antibody Working Solution	5.5 mL
5	Substrate Reagent A	6 mL
6	Substrate Reagent B	6 mL
7	Stop Solution	6 mL
8	10×Reconstitution Buffer	50 mL
9	20×Concentrated Wash Buffer	40 mL
10	Plate Sealer	3 pieces
11	Sealed Bag	1 piece
12	Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

### Additional materials required:

#### Other materials required but not supplied

- **Instrument:** Microplate reader, Printer, Homogenizer, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).
- **High-precision transferpettor:** Single channel (20-200 µL, 100-1000 µL), Multichannel (300 µL).
- **Reagent:** Methanol、N-hexane

## 5. Experimental Preparation

Bring all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

### 1. Sample pre-treatment Notice:

Experimental apparatus should be clean, and use disposable pipette tips to avoid cross-contamination during the experiment.

### 2. Solution preparation

*Please prepare solution according to the number of samples. Don't use up all components at once!*

Solution 1: 70% Methanol

**Methanol** (V): Deionized water (V) =7: 3.

Solution 2: Reconstitution Buffer

Dilute the **10×Reconstitution Buffer** with deionized water (1:9). Store at 4°C for a month.

Solution 3: Wash Buffer

Dilute the **20×Concentrated Wash Buffer** with deionized water.

(20×Concentrated Wash Buffer (V): Deionized water (V) =1:19)

### 3. Sample pre-treatment procedure

*Targets may be distributed unevenly, resulting in no detection. To avoid this, ensure to take sufficient samples when sampling.*

#### 3.1 Pre-treatment of corn, feed sample:

1. Weigh 1g of crushed homogenate into the 50 mL centrifuge tube, add 5 mL of deionized water, vortex for 5 min. Centrifuge at 4000 r/min for 10 min at room temperature;
2. Take 0.1 mL of supernatant, add 0.9 mL of **Reconstitution Buffer** (Solution 2), vortex for 2 min;
3. Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 50, detection limit: 150 ppb**

#### 3.2 Pre-treatment of edible oil sample:

1. Take 5 mL of sample into the 50 mL centrifuge tube, add 8 mL of **N-hexane** and 5 mL of **70% Methanol** (Solution 1), vortex for 5 min, centrifuge at 4000 r/min for 10 min at room temperature.
2. Discard the supernatant, take 100  $\mu$ L of under layer liquid, and add 1.9 mL of **Reconstitution Buffer** (Solution 2), mix fully.
3. Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 20, detection limit: 20 ppb**

## 6. Assay Procedure

Bring all reagents and samples to room temperature (25°C) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2~8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples must be tested in duplicate.**
2. **Add sample:** add 50  $\mu$ L of **Standard or Sample** per well, then add 50  $\mu$ L of **HRP Conjugate** to each well, then add 50  $\mu$ L of **Antibody Working Solution**, cover the plate with plate sealer, vortex for 5s gently to mix thoroughly. Incubate at 25°C for 30 min away from direct sunlight.
3. **Wash:** uncover the sealer carefully, remove the liquid. Immediately add 300  $\mu$ L of **Wash Buffer** (Solution 3) to each well and wash. Repeat wash procedure for 5 times, 30 s intervals/time. Invert the plate and pat it against absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Colour Development:** add 50  $\mu$ L of **Substrate Reagent A** to each well, and then add 50  $\mu$ L of **Substrate Reagent B**. Gently vortex for 5s to mix thoroughly. Incubate at 25°C for 15min away from direct sunlight (The reaction time can be extended according to the actual colour change).
5. **Stop reaction:** add 50  $\mu$ L of **Stop Solution** to each well, vortex gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

## 7. Data Analysis

### 1. Absorbance (%) = $A/A_0 \times 100\%$

A: Average absorbance of standard or samples  $A_0$ :

Average absorbance of 0 ppb Standard

### 2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

## 8. Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. ELISA Microplate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the FSES0079. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other FSES0079 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue colour. When OD value of standard (concentration: 0) < 0.5 unit (A450nm < 0.5), it indicates the reagents are deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

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### **Contact Details**



Email: [info@AsssayGenie.com](mailto:info@AsssayGenie.com)

Web: [www.AssayGenie.com](http://www.AssayGenie.com)