



Technical Manual

α -Amylase Activity Assay Kit

- **Catalogue Code: MAES0020**
- **Size: 96T**
- **Research Use Only**

1. Key features and Sample Types

Detection method:

Colorimetric method

Specification:

96T

Range:

0.97-34.74 U/g tissue

Sensitivity:

0.97 U/g tissue

Storage:

2-8°C for 6 months

Expiry:

See Kit Label

Experiment Notes:

This kit is for **research use only**.

Instructions should be strictly followed. Changes of operation may result in unreliable results.

The validity of kit is 6 months.

Do not use components from different batches of kit.

2. Background

Amylase is an enzyme that catalyses the hydrolysis of starch into sugars. Amylase is present in the saliva of humans and some other mammals, where it begins the chemical process of digestion. Foods that contain large amounts of starch but little sugar, such as rice and potatoes, may acquire a slightly sweet taste as they are chewed because amylase degrades some of their starch into sugar. The pancreas and salivary gland make amylase (α -amylase) to hydrolyse dietary starch into disaccharides and trisaccharides which are converted by other enzymes to glucose to supply the body with energy. Plants and some bacteria also produce amylase. As diastase, amylase was the first enzyme to be discovered and isolated (by Anselme Payen in 1833). Specific amylase proteins are designated by different Greek letters. All amylases are glycoside hydrolases and act on α -1,4-glycosidic bonds.

3. Intended Use

This kit can measure β -Amylase activity in plant and animal tissue samples.

4. Detection Principle

The reducing sugar reacts with 3,5-dinitrosalicylic acid under heating conditions to produce a brown-red substance, which is inactivated by the thermolabile nature of β -amylase, and then the enzyme activity of α -amylase is determined.

5. Kit components & storage

Item	Specification	Storage
Substrate	10 mLx1 vial	2-8°C, 6 months
Chromogenic Agent	20 mLx1 vial	2-8°C, 6 months, avoid direct sunlight
Standard (10 mg/mL)	1.5 mLx1 vial	2-8°C, 6 months
Microplate	96 wells	No requirement
Plate Sealer	2 pieces	

Materials required but not supplied

- Micropipettor
- 95°C Water bath
- Centrifuge
- Microplate Reader (535-540 nm)
- Tips (10 μ L, 200 μ L, 1000 μ L)
- EP tubes (1.5 mL, 2 mL)
- Double distilled water

6. Assay Notes:

1. For measuring the OD value, if there is precipitation, centrifuge at 4000 g for 5 min at room temperature and take the supernatant for determination.
2. When the absolute OD value is greater than 0.747, sample should be diluted appropriately.

7. Reagent Preparation

1. Bring all reagents to room temperature before use. Before the experiment, preheat substrate and chromogenic agent at 40°C for 10 min.
2. If there is precipitation in substrate, please use it after heating and dissolving at 70°C.
3. If there is yellow precipitation in chromogenic agent, please use it after heating and dissolving at 70°C.

8. Sample Preparation

Tissue sample:

1. Weigh 0.1 g sample, add 0.9 mL of distilled water and homogenized with a homogenizer, then transfer to the EP tube, incubate at room temperature for 15 min and oscillate every 5 min. Centrifuge at 3000 g at room temperature for 10 min, take the supernatant and add double distilled water to the final volume of 10 mL, mix fully and it is the prepared sample.

Sample Notes:

The concentration should be determined before performing the assay. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.

If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.

Dilution of Samples:

Large variances in results may be seen when performing pre-experiments. Dilute the sample according to the result of the pre-experiment and the detection range (0.97-34.74 U/g tissue).

The recommended dilution factor for different samples is as follows (for reference only).

Sample Type:	Dilution Factor
1% Epipremnum aureum tissue homogenate	1
1% Green pepper tissue homogenate	1
1% Corn grain tissue homogenate	1
1% Daucus carota tissue homogenate	1

Note: The diluent is double distilled water.

9. Assay Protocol

Ambient Temperature: 25-30°C

Optimum detection wavelength: 540 nm

Plate Set Up:

	1	2	3	4	5	6	7	8	9	10	11	12
A	A	A	S1'	S1	S9'	S9	S17'	S17	S25'	S25	S33'	S33
B	B	B	S2'	S2	S10'	S10	S18'	S18	S26'	S26	S34'	S34
C	C	C	S3'	S3	S11'	S11	S19'	S19	S27'	S27	S35'	S35
D	D	D	S4'	S4	S12'	S12	S20'	S20	S28'	S28	S36'	S36
E	E	E	S5'	S5	S13'	S13	S21'	S21	S29'	S29	S37'	S37
F	F	F	S6'	S6	S14'	S14	S22'	S22	S30'	S30	S38'	S38
G	G	G	S7'	S7	S15'	S15	S23'	S23	S31'	S31	S39'	S39
H	H	H	S8'	S8	S16'	S16	S24'	S24	S32'	S32	S40'	S40

Note: A-H, standard wells; S1'-S40', control wells; S1-S40, sample wells

10. Operation Steps

The preparation of standard curve

Dilute standard (10 mg/mL) with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4 mg/mL.

The measurement of standard

1. Take 1.5 mL EP tube and number the tubes from A to H in duplication, add 75 μ L of standard solution with different concentrations to the corresponding tubes.
2. Add 75 μ L of substrate to each tube.
3. Add 150 μ L of chromogenic agent to each tube.
4. Mix fully and incubate at 95°C for 5 min. Cool the tubes with running water and take 250 μ L of supernatant to the microplate. Measure the OD value of each well with microplate reader at 540 nm.

The measurement of sample

- Sample tube:** Add 75 μL of sample to the corresponding tubes.
Control tube: Add 75 μL of sample to the corresponding tubes.
- Incubate at 70°C water bath for 15 min and cool the tubes with running water.
- Sample tube:** Add 75 μL of substrate to the corresponding tubes.
Control tube: Add 75 μL of double distilled water to the corresponding tubes.
- Incubate the sample tubes and control tubes at 40°C water bath for 5 min.
- Add 150 μL of chromogenic agent to each tube.
- Mix fully and incubate at 95°C for 5 min. Cool the tubes with running water and take 250 μL of supernatant to the microplate. Measure the OD value of each well with microplate reader at 540 nm.

Operation Table

The measurement of standard

	Standard tubes
Standard solution with different concentrations (μL)	75
Substrate (μL)	75
Chromogenic agent (μL)	150
Mix fully and incubate at 95°C for 5 min. Cool the tubes with running water and take 250 μL of supernatant to the microplate. Measure the OD value of each well with microplate reader at 540 nm.	

The measurement of sample

	Control tubes	Sample tubes
sample (μL)	75	75
Incubate at 70°C water bath for 15 min and cool the tubes with running water.		
Double distilled water (μL)	75	
Substrate (μL)		75
Incubate the sample tubes and control tubes at 40°C water bath for 5 min.		
Chromogenic agent (μL)	150	150
Mix fully and incubate at 95°C for 5 min. Cool the tubes with running water and take 250 μL of supernatant to the microplate. Measure the OD value of each well with microplate reader at 540 nm.		

11. Calculations

Plot the standard curve by using OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the OD value of sample. The standard curve is: $y = ax + b$.

1. Calculate according to the protein concentration of the sample

Definition: The production of 1 mg reducing sugar catalyzed by 1 mg of tissue protein per minute that is defined as an enzyme activity unit.

$$\alpha\text{-amylase activity (U/mg prot)} = (\Delta A - b) \div a \times V_3 \div t \div V_2 \div C_{pr}$$

2. Calculate according to the fresh weight of sample

Definition: The production of 1 mg reducing sugar catalyzed by 1 g of tissue per minute that is defined as an enzyme activity unit.

$$\alpha\text{-amylase activity (U/g fresh weight)} = (\Delta A - b) \div a \times V_3 \div t \div w \times \frac{V_1}{V_2} \times f$$

y: $OD_{Standard} - OD_{Blank}$. (OD_{Blank} is the OD value when the standard concentration is 0)

x: The concentration of standard.

a: The slope of standard curve.

b: The intercept of standard curve.

f: Dilution factor of sample before tested.

ΔA : $OD_{Sample} - OD_{Control}$

V_1 : The volume of prepared tissue sample in sample preparation step (10 mL).

V_2 : The volume of sample added to the reaction (0.075 mL).

V_3 : The volume of enzymatic reaction (the volume of sample + the volume of substrate = 0.15 mL).

t: The time of enzymatic reaction (5 min).

w: The weight of tissue sample (0.1 g).

C_{pr} : Concentration of protein in sample (mgprot/mL).

12. Performance Characteristics

Detection Range	0.97-34.74 U/g tissue
Sensitivity	0.97 U/g tissue
Average recovery rate (%)	98
Average inter-assay CV (%)	3.9
Average intra-assay CV (%)	3.0

Analysis

Take 0.1 g of green pepper, treat the sample according to the sample preparation step, carry the assay according to the operation table.

The results are as follows:

$y = 0.8729x - 0.0112$, the average OD value of the sample is 0.368, the average OD value of the control is 0.247, and the calculation result of α -amylase activity is:

α -amylase activity (U/g fresh weight)

$$\begin{aligned} &= (0.368 - 0.247 + 0.0112) \div 0.8729 \times 0.15 \div 5 \div 0.1 \times 10 \div 0.075 \\ &= 6.06 \text{ U/g tissue} \end{aligned}$$

Safety Notes

Some of the reagents in the kit contain dangerous substances. Prevent touching skin and clothing.

Wash immediately with plenty of water if touching it carelessly.

All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

Before the experiment, read the instructions carefully, and wear gloves and work clothes.

Notes:

Notes:

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Contact Details



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