

Technical Manual

Alkaline Phosphatase (ALP) Colorimetric Assay Kit (PNPP method)

- Catalogue Code: MAES0023
- Size: 96T
- Research Use Only

1. Key features and Sample Types

Detection method:

Colorimetric method

Specification:

96T

Range:

0.27-50.8 U/L

Sensitivity:

0.27 U/L

Storage:

-20°C for 6 months

Expiry:

See Kit Label

Experiment Notes:

This kit is for **research use only.**

Instructions should be strictly followed. Changes of operation may result in unreliable results.

The validity of kit is 6 months.

Do not use components from different batches of kit.

2. Background

Alkaline phosphatase (ALP) is a group of cytomembrane-related enzymes with hydrolysis and transfer activity, acting on a variety of phosphate substrates. ALP is a homologous dimerase and each catalytic site contains three metal ions. There are four isozymes in humans: tissue nonspecific ALP, intestinal ALP, placental ALP and genital cell ALP.

3. Intended Use

This kit can be used to measure alkaline phosphatase (ALP) activity in animal tissue, serum (plasma) and cell samples.

4. Detection Principle

Under alkaline conditions, alkaline phosphatase catalyzes the hydrolysis of pnitrobenzene phosphate disodium to produce p-nitrophenol and phosphoric acid. Under strong alkaline conditions, p-nitrophenol is bright yellow and has a maximum absorption peak at 405 nm. Therefore, the activity of ALP can be calculated by measuring the OD value at 405 nm.

5. Kit components & storage

ltem	Specification	Storage
Buffer Solution	60 mL×1 vial	-20°C, 6 months
Substrate	2 vials Lyophilized	-20°C, 6 months, away from direct sunlight
p-Nitrophenol Standard Solution (10 mmol/L)	0.4 mL×1 vial	-20°C, 6 months, away from direct sunlight
Stop Solution	12 mL × 1 vial	-20°C, 6 months
Microplate	96 wells	No requirement
Plate Sealer	2 pieces	

Materials required but not supplied

- Micropipettor
- Incubator
- Centrifuge
- Microplate Reader (400-415 nm)
- Tips (10 μL, 200 μL, 1000 μL)
- EP tubes (1.5 mL, 2 mL)
- Normal Saline (0.9% NaCl)

6. Assay Notes:

Substrate working solution and standard should be stored away from direct sunlight.

7. Reagent preparation:

- 1. Bring all reagents to room temperature before use.
- 2. Preparation of **substrate working solution**: Dissolve substrate with 3 mL of buffer solution. The prepared solution can be stored at -20°C with shading light for 24 hours.
- 3. Preparation of **500 µmol/L standard:** Mix the 10 mmol/L standard and buffer solution at the ratio of 1:19. Prepare the fresh needed amount before use and store away from direct sunlight for detection.

8. Sample Preparation

- 1. Serum (Plasma): Detect the sample directly.
- Tissue Sample: Weigh the tissue. Add normal saline (0.9% NaCl) in a weight (g): volume (mL) ratio of 1:9. Homogenize mechanically in an ice bath to fully break the cells. Centrifuge at 10000 g for 10 min at 4°C and collect the supernatant for measurement. Meanwhile, determine the protein concentration of the supernatant.
- 3. **Cell Sample:** Add normal saline (0.9% NaCl) at a ratio of cell number $(3*10^8)$: 0.9% NaCl (μ L) = 1:200-400.
- 4. **Sample Requirements:** The Sample Solution should not include inhibitors of alkaline phosphatase, such as EDTA, fluoride ion and citrate.

Sample Notes:

The concentration should be determined before preforming the assay. If the sample concentration is not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.

If the sample type is not included in the manual, a preliminary experiment is suggested to verify the validity.

Dilution of Samples:

Large variances in results may be seen when performing pre-experiments. Dilute the sample according to the result of the pre-experiment and the detection range (0.27-50.8 U/L).

The recommended dilution factor for different samples is as follows (for reference only).

Sample Type:	Dilution Factor
Human serum	10-20
Mouse serum	20-30
Rat serum	10-20
Mouse plasma	10-20
HepG2 supernatant	8-12
10% Rat kidney tissue homogenate	400-1200
10% Mouse liver tissue homogenate	10-15
10% Rat lung tissue homogenate	50-100
10% Mouse brain tissue homogenate	30-50
Human urine	1
Note: The diluent is buffer solution	

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9. Assay Protocol

Ambient Temperature: 25-30°C

Optimum detection wavelength: 405 nm

Plate Set Up:

	1	2	3	4	5	6	7	8	9	10	11	12
Α	А	А	S0	S8	S16	S24	S32	S40	S48	S56	S64	S72
В	В	В	S1	S9	S17	S25	S33	S41	S49	S57	S65	S73
С	С	С	S2	S10	S18	S26	S34	S42	S50	S58	S66	S74
D	D	D	S3	S11	S19	S27	S35	S43	S51	S59	S67	S75
Е	E	E	S4	S12	S20	S28	S36	S44	S52	S60	S68	S76
F	F	F	S5	S13	S21	S29	S37	S45	S53	S61	S69	S77
G	G	G	S6	S14	S22	S30	S38	S46	S54	S62	S70	S78
н	н	Н	S7	S15	S23	S31	S39	S47	S55	S63	S71	S79

Note: A-H, standard wells; S0, control wells; S1-S79, sample wells.

10. Operation Steps

The preparation of standard curve

Dilute 500 µmol/L standard solution with buffer solution to a serial concentration. The recommended dilution gradient is as follows: 500, 400, 320, 240, 160, 80, 40, 0 µmol/L.

The measurement of samples

- Standard well: Add 50 μL of standard solution with different concentrations to the wells.
 Sample well: Add 50 μL of sample to the wells.
 Control well: Add 50 μL of buffer solution into the control wells.
- 2. Add 50 μ L of substrate working solution into the control wells and sample wells. Add 50 μ L of buffer solution into the standard wells.
- 3. Incubate at 37°C for 10 min.
- 4. Add 100 µL of stop solution into each well.
- 5. Mix fully with microplate reader for 5 s. Measure the OD values of each well at 405 nm with microplate reader.

Operation Table

	Standard well	Sample well	Control well		
Standards with different concentrations (µL)	50				
Sample (μL)		50			
Buffer solution (μL)	50		50		
Substrate working solution (µL)		50	50		
Incubate at 37°C for 10 min.					
Stop solution (µL)	100	100	100		
Mix fully with microplate reader for 5 s with microplate reader.	. Measure the OD	values of each	well at 405 nm		

11. Calculations

Plot the standard curve by using OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the OD value of sample. The standard curve is: y = ax + b.

1. Serum (plasma) sample:

Definition: The amount of 1 µmol p-nitrophenol produced by 1 L serum (plasma) per minute catalyze the substrate at 37°C is defined as 1 activity unit.

$$\frac{ALP \text{ activity}}{(U/L)} = (\Delta A-b) \div a \div T \times f$$

2. Tissue and cells sample:

Definition: The amount of 1 µmol p-nitrophenol produced by 1 g sample protein per minute catalyze the substrate at 37°C is defined as 1 activity unit.

 $\begin{array}{l} ALP \ activity\\ (U/gprot) \end{array} = (\Delta A - b) \div a \div T \times f \div C_{pr} \end{array}$

y: $OD_{Standard} - OD_{Blank}$ (OD_{Blank} is the OD value when the standard concentration is 0).

x: The concentration of standard.

a: The slope of standard curve .

b: The intercept of standard curve.

ΔA: OD_{Sample} – OD_{Control}

 $\ensuremath{\textbf{f}}$: Dilution factor of sample before test

T: Reaction time, 10 min

 C_{pr} : Concentration of protein in sample, gprot/L.

12. Performance Characteristics

Detection Range	0.27-50.8 U/L
Sensitivity	0.27 U/L
Average inter-assay CV (%)	4.6
Average intra-assay CV (%)	1.2

Analysis

For human serum, dilute human serum with buffer solution for 10 times, take 50 μ L of diluted samples, carry the assay according to the operation table. The results are as follow.

The results are as follows:

standard curve: y = 0.0027 x - 0.0051, the OD value of the control is 0.099, the OD value of the sample is 0.544, and the calculation result is:

ALP activity
$$\left(\frac{U}{L}\right) = (0.544 - 0.099 + 0.0051) \div 0.0027 \div 10 \times 10$$

= 166.70 U/L

Safety Notes

Some of the reagents in the kit contain dangerous substances. Avoid touching skin and clothing.

Wash immediately with plenty of water if touching it carelessly.

All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

Before the experiment, read the instructions carefully, and wear gloves and work clothes.

Notes:

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Contact Details



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