TECHNICAL MANUAL

FMC63 scFv ELISA Kit

- **SKU CODE:** GXEL00534
- **SIZE:** 96 Wells
- **DETECTION PRINCIPLE:** Sandwich ELISA
- **RUO:** Research-Use-Only

Revised: 10/26

FMC63 scFv ELISA Kit

Please read entire manual carefully before starting experiment.

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1. Key Features

Sample type

Cell lysates, tissue lysates, plasma, and serum

Sensitivity

The minimum detectable dose (MDD) is reported in the COA included with each kit.

Range

Provided in the Certificate of Analysis (COA) included with the product.

Detection method

Colorimetric (450 nm)

Assay type

Quantitative Sandwich ELISA

2. Storage & Expiry Date

Assay Genie ELISA Kits are shipped on ice packs. Please store this ELISA Kit and/or components as described below. Date of expiration is on the ELISA Box label.

1. For Unopened Kits:

Store the entire, unopened kit at 2-8°C in its original box with a shelf life of 6 months.

For optimal performance extending up to 1 year, it is recommended to store components separately:

- **-20°C:** Standard, Detection Reagent A, Detection Reagent B, and Pre-coated Microplate.
- **2-8°C:** Standard Diluent, Assay Diluent, 20X Wash Buffer, Color Reagent, Stop Solution, and Plate Sealers.

2. For Opened/Used Kits:

- Return any unused well strips to the foil pouch with the desiccant and reseal it tightly.
- Store all remaining reagents according to the conditions above (separated at -20°C or 2-8°C as applicable).
- Reagents are typically stable for about 1 month after the kit is first opened.

Note: Please always refer to the specific instructions provided with your kit.

3. Product Description

This kit is based on a quantitative sandwich enzyme-linked immunosorbent assay (ELISA). Standards and samples are added to the pre-coated microplate wells, where FMC63 scFv participates in a format-specific immunoreaction. After the required incubation and washing steps, enzyme-conjugated reagent and substrate solution are added sequentially to generate a measurable colorimetric signal.

The optical density (OD) value is positively correlated with the concentration of FMC63 scFv in the sample. The concentration of FMC63 scFv is calculated by interpolation from the standard curve.

This dual function kit includes validated Bradford Reagent to quantify total protein concentration for accurate sample normalization.

4. Kit Contents

Part	Format	Description	Storage Conditions
1	1 plate	Pre-coated Microplate	Store sealed at -20°C.
2	2 bottles	FMC63 scFv Standard	Store at -20°C.
3	1 vial	Detection A	Store at -20°C.
4	1 vial	Detection B	Store at -20°C.
5	1 bottle	Standard Diluent	Store at 4°C.
6	1 bottle	Assay Diluent	Store at 4°C.
7	1 bottle	20 × Wash Buffer	Store at 4°C.
8	1 bottle	Color Reagent	Store at 4°C.
9	1 bottle	Stop Solution	Store at 4°C.
10	1 vial	Bradford Reagent	Store at RT
11	4 strips	Plate Sealers	Store at RT.

* Provided this is within the expiration date of the kit.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm, with the correction wavelength set at 630 nm or 620 nm.
- Pipettes and pipette tips.
- Deionized or distilled water.
- 500 mL graduated cylinder.
- Squirt bottle, manifold dispenser, or automated microplate washer.
- Test tubes for dilution of standards.

5. Precautions

1. This kit is for research purposes only and not for diagnostics or therapeutic uses.
2. Store all components as listed in this manual. Do not use the ELISA Kit after its expiration date.
3. Allow all reagents and samples to reach room temperature before use.
4. Ensure unopened and unused plates are kept dry to avoid contamination.
5. Before using the kit, centrifuge tubes to spin down standard and/or antibody.
6. Prepare all reagents, samples and standards as directed in this manual.
7. Duplicate wells are recommended for both standard and sample testing.
8. Do not let the microplate wells dry during the assay.
9. Maintain consistent incubation times and temperatures as variations can affect results.
10. Do not reuse tips and tubes to avoid cross contamination.
11. Avoid using the reagents from different batches together.

6. Sample Collection & Storage

***Note**: For information regarding validation data in specific samples, please contact our Technical Support Team at techsupport@assaygenie.com.*

General Considerations

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

Handle all blood and serum as if capable of transmitting infectious agents. The NCCLS provides recommendations for handling and storing cell lysates, tissue lysates, plasma, and serum specimens (Approved Standard-Procedures for the Handling and Processing of Blood Specimens, H18-A. 1990).

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.

Other Sample Types

For more information about how to process other sample types, (e.g., body fluids, breast milk & more), please contact our Tech Support Team at techsupport@assaygenie.com.

6.1. Protein Quantification (Optional)

To quantify total protein levels, use the Bradford Reagent included in this kit. Visit Bradford Protein Assay Protocol to view the full protocol.

7. Reagent Preparation

Bring all reagents to room temperature before use.

20-fold Wash Buffer Concentrate - If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 25 mL of Wash Buffer Concentrate to 475 mL of deionized or distilled water to prepare 500 mL of **Wash Buffer**.

Cell lysates, tissue lysates, plasma, and serum - Serum and plasma samples require a 10-fold dilution. A suggested 10-fold dilution is 10 µL of sample + 90 µL of Standard Diluent (diluted 1:9). If the sample value is outside the range of the standard curve, the dilution can be adjusted appropriately and the assay can be redetermined. If the antibody concentration in the sample can be estimated and the assay can be performed simultaneously by diluting several gradients prior to the experiment.

Standard - Reconstitute with 1 mL **Standard Diluent**, this reconstitution produces a **stock solution**. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. The appropriate **Standard Diluent** serves as the zero standard (0 ng/mL).

Detection A (working solution) - Shake and mix before used. Centrifuge instantaneously with palm centrifuge to make the liquid at the bottom of the tube. Dilute the **Detection A** 1: 100 times to the working concentration with **Assay Diluent**.

Detection B (working solution) - Shake and mix before used. Centrifuge instantaneously with palm centrifuge to make the liquid at the bottom of the tube. Dilute the **Detection B** 1: 100 times to the working concentration with **Assay Diluent**.

8. Assay Procedure

Bring all reagents and samples to room temperature before use. It is recommended that all standards and samples be assayed in duplicate.

1. Prepare all reagents and working standards as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal.
3. Add 100 µL standards or samples to each well. Cover with the adhesive strip provided. Incubate for 1 hour at 37°C.
4. Aspirate each well and wash, repeating the process three times. Wash by filling each well with Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
5. Add 100 µL of Detection A working solution to each well. Cover with a new adhesive strip. Incubate for 1 hour at 37°C.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 µL of Detection B working solution to each well. Cover with a new adhesive strip. Incubate for 30 minutes at 37°C.
8. Aspirate each well and wash, repeating the process five times. Wash by filling each well with Wash Buffer (300 µL) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
9. Add 100 µL of Color Reagent to each well. Incubate for 15 minutes at 37°C. Protect from light.
10. Add 50 µL of Stop Solution to each well. The color in the wells should change from blue to yellow. If the color in the wells is green or the color change does not appear uniform, gently tap the plate to ensure thorough mixing.
11. Determine the optical density of each well within 10 minutes, using a microplate reader set to 450 nm. If wavelength correction is available, set to 630 nm or 620 nm. If wavelength correction is not available, subtract readings at 630 nm or 620 nm from the readings at 450 nm. This subtraction will correct for optical imperfections in the plate. Readings made directly at 450 nm without correction may be higher and less accurate.

9. Calculation Results

Average the duplicate readings for each standard and sample. Construct a standard curve by plotting the mean absorbance for each standard on the Y-axis against the concentration on the X-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the FMC63 scFv concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data.

If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.

10. Typical Data

Sensitivity

The minimum detectable dose (MDD) for FMC63 scFv is reported in the COA included with each kit.

Precision

- **Intra-Assay Precision (Precision within an assay):** <15%
  Three samples of known concentration were tested sixteen times on one plate to assess intra-assay precision.
- **Inter-Assay Precision (Precision between assays):** <15%
  Three samples of known concentration were tested in twenty four separate assays to assess inter-assay precision.

Stability

When the kit was stored at the recommended temperature for 12 months, the signal intensity decreased by less than 30%.

For unopened kits, if you want to prolong the storage time, please store the Standard, Detection A, Detection B and Microplate at -20 °C, the rest reagents should be store at 4°C.

11. Troubleshooting Guide

Problem	Probable Cause	Solution
Poor Precision	Wells are not washed or aspirated properly	Make sure the wash apparatus works properly and wells are dry after aspiration
	Bubbles in the wells	Tap plate gently to disperse bubbles
	Wells are scratched with pipette tip or washing needles	Dispense and aspirate solution into and out of wells with caution
	Particulates are found in the samples	Remove any particulates by centrifugation prior to the assay
High background	Plate is not washed properly	Make sure the wash apparatus works properly
	Incorrect incubation times and/or temperatures	The OD value increased gradually along with the time. Reduce the color developing time properly
Weak/No Signal	Pipetting errors	Make sure the pipette is calibrated
	The working solution not be prepared immediately before use	The working solution should be prepared immediately before use and should not be stored
	Volumes errors	Repeat assay with the required volumes in manual
	The plate is not incubated for proper time or temperature	Follow the manual to repeat assay
	Detection A working solution is not completely mixed with the samples	After adding the Detection A into the wells, make sure the detection A and the samples are mixed thoroughly

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Assay Genie 100% money-back guarantee!

If you are not satisfied with the quality of our products and our technical team cannot resolve your problem, we will give you 100% of your money back.

Manufacturers Statement: This final kit system is assembled and quality-released by Assay Genie Limited.